Separation and analysis of poly(lactide-co-glycolide) in Trelstar® 22.5 mg formulation

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Introduction

Poly(lactide-co-glycolide) (PLGA) has been used in most of the injectable, long-acting formulations approved by the U.S. Food and Drug Administration (FDA). Some formulations utilized complex mixtures of different types of PLGA to achieve specific release profiles. A proposed generic formulation needs to match the reference listed drug for qualitative and quantitative (Q1/Q2) sameness of the PLGA, including polymer mixtures and chirality of lactide unit. The purpose of this study is to leverage the lactoselectivity effect of various solvents to separate and analyze the components of Trelstar® 22.5 mg formulation. In addition to the previously established parameters (L:G ratio, endcap, blockiness, molecular weight, branching) samples were also assayed by an advanced proton nuclear magnetic resonance (HNMR) technique to determine chirality of the lactide component.

Methods

Samples of Trelstar[®] 22.5 mg (Allergan) were extracted to obtain PLGA as previously described [1]. Briefly, the contents of each vial were loaded into centrifuge tubes. Dichloromethane (DCM) was added to each tube and dissolved with orbital agitation at 100 RPM/30°C overnight. The supernatant was passed through a syringe filter into hexane to precipitate out the PLGA extract [1]. Afterwards, semi-solvent extraction was performed according to previously established methods for PLGA separation by lactide content [2]. Briefly, each semi-solvent was added to generate a total concentration of 25 mg/ml. The tube was incubated 30°C overnight. The solution was centrifuged and the soluble supernatant portion removed. The remaining material was dried and the process was repeated. This was done using xylenes, isopentyl-acetate, toluene, butyl-acetate, 2-pentanone, and butanone, in that order, to separate out PLGA from high lactide to low lactide fraction. Analysis of each fraction was performed as previously described [3]. Briefly, lactide content was determined by HNMR comparison of lactide peak (1H, 5.2 ppm) and glycolide peak (2H, 4.8 ppm). Blockiness (Rc) were determined by C13NMR by dividing GA-LA peak (166.4 ppm) from the GA-GA peak (166.3 ppm). Molecular weight of each fraction was determined by GPC-4D utilizing universal calibration. Prior to chirality analysis, samples were re-dissolved in DCM, passed through a 0.45 um polyvinylidene fluoride (PVDF) filter and precipitated in methanol followed by vacuum drying. Chirality analysis was performed according to previous methods [4]. Briefly, data were obtained using a Bruker Avance DRX-500 NMR spectrometer running TopSpin 1.3 software and equipped with a 5mm TBI Z-gradient probe. The sample temperature was set to ca. 299K. Data were acquired with homonuclear decoupling (Bruker pulse sequence zghd) of the methyl-group resonances using the following parameters: observe pulse width ca. 90 degrees, acquisition time 4.4 seconds, relaxation delay 1 second, 16 - 64 scans averaged following 4 dummy scans.

Methods (cont.)

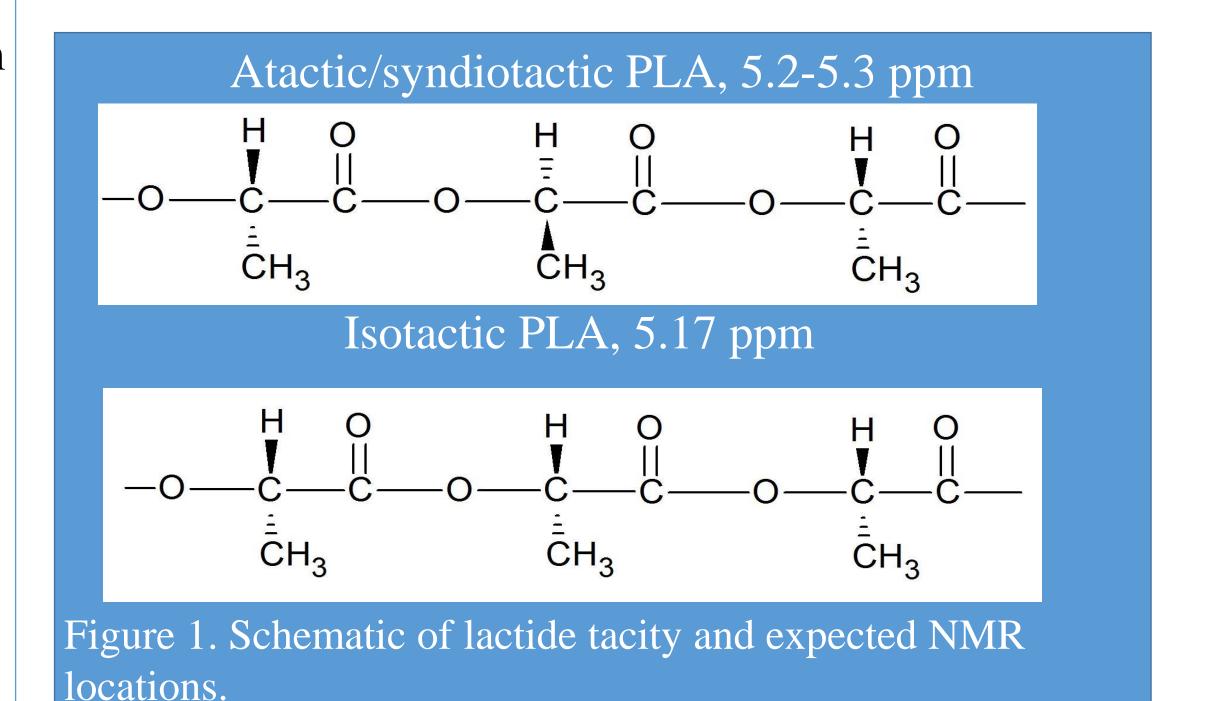
The specific decoupling frequency (Bruker parameter O2) was set by inspection for each sample after obtaining the conventional (non-decoupled) spectrum. Raw data were processed with exponential multiplication (line broadening factor 0.5) prior to Fourier transformation without zero-filling. Both known samples of PLGA (DL or L) and Trelstar fractions were tested.

Results

Two Trelstar® batches (lot# 16-008386 and lot #18-01012) were assayed and the resultant data are shown in **Table 1.**

Table 1. Trelstar® Fraction Separation Analysis (Average ± STDEV, N=2 lots)					
Solvent (fraction)	Percent polymer (w/w%)	Lactide content (%L, NMR)	Mw (GPC-4D)	Mn (GPC-4D)	Rc (NMR)
Original Mixture	100%	76.9 ± 0.1	$41,377 \pm 135$	$31,475 \pm 81$	0.78 ± 0.01
Xylenes	6.0 ± 0.1	84.0 ± 0.1	$13,063 \pm 2695$	8755 ± 4799	0.46 ± 0.16
Isopentyl acetate	15.8 ± 0.8	82.8 ± 0.1	$24,653 \pm 1316$	$19,429 \pm 811$	0.48 ± 0.08
Toluene	25.5 ± 1.3	82.9 ± 0.1	$47,790 \pm 939$	39,084± 2588	0.55 ± 0.12
Butyl acetate	12.6 ± 0.2	74.2 ± 0.2	$26,592 \pm 665$	$22,760 \pm 99$	0.81 ± 0.02
2-Pentanone	14.7 ± 0.2	72.5 ± 0.2	$35,483 \pm 264$	$29,658 \pm 88$	0.88 ± 0.001
Butanone	24.7 ± 0.7	70.8 ± 0.2	$52,930 \pm 640$	45,267 ± 1467	0.89 ± 0.01
Butanone residual	0.6 ± 0.4	70.5 ± 0.6	NT*	NT*	NT*
* NT = Not Tested, too little quantity extracted to test.					

Analysis was performed to determine if parameters relevant to the chirality nature of the polymers could be defined. In PLGA's >75L a pronounced peak based on isotactic lactide at ~ 5.17 ppm (**Figures 1-3**) could be used to assay chirality.



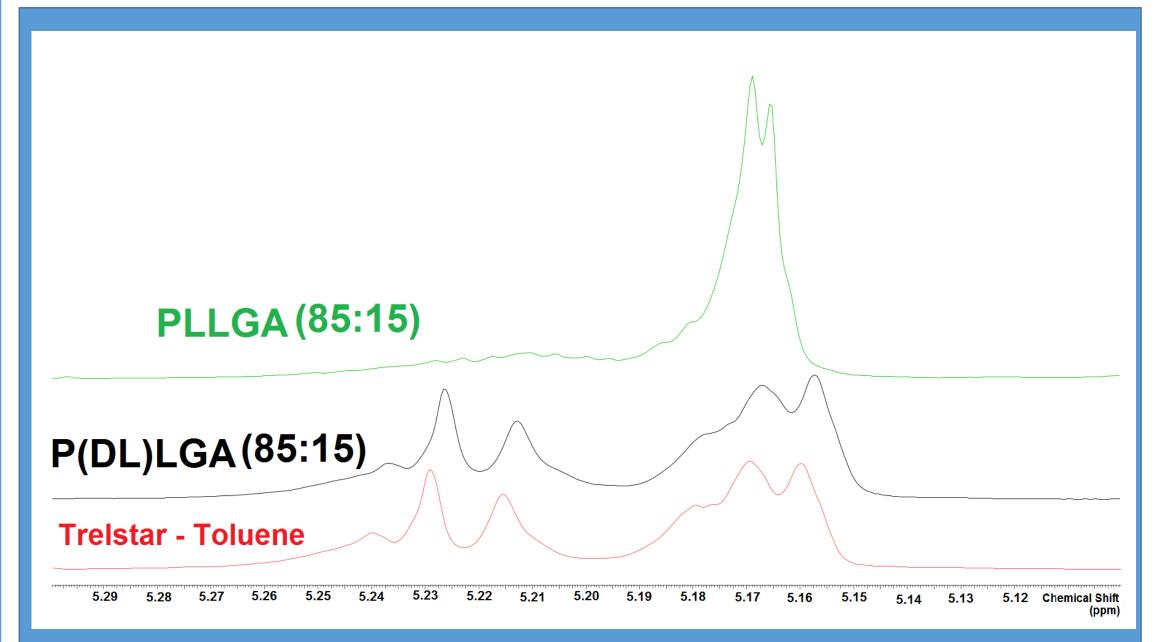


Figure 2. NMR comparison of Trelstar Toluene fraction with PLLGA and P(DL)LGA standards (85:15 LA:GA).

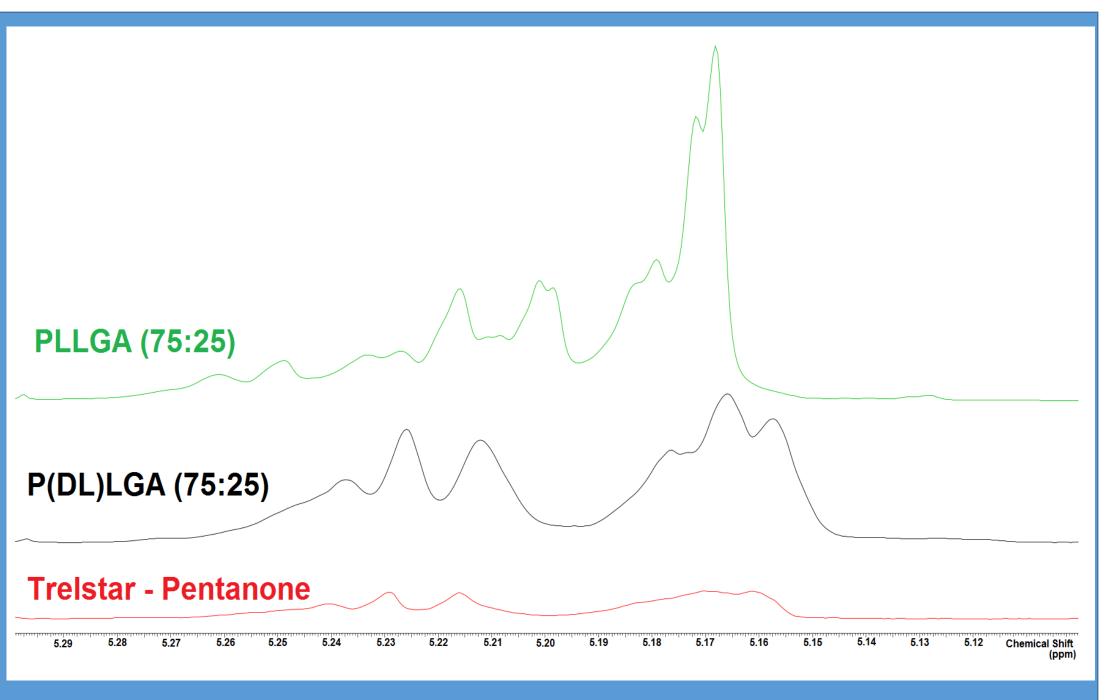


Figure 3. NMR comparison of Trelstar Pentanone fraction with PLLGA and P(DL)LGA standards (75:25 LA:GA).

Conclusion

Without fractionation, analytical techniques (such as NMR) only provide the average values of the fractions of PLGA's present in a polymer sample. Using semi-solvents, Trelstar® 22.5 mg formulation was successfully separated into fractions based on lactide content of PLGA and each component assayed individually. In addition to conventional parameters, the chirality of high-lactide content PLGA's can be obtained by decoupled NMR analysis and Trelstar fractions contain 'DL' PLGA.

References

- [1] J. Garner, S. Skidmore, H. Park, K. Park, S. Choi, and Y. Wang: A protocol for assay of poly(lactide-coglycolide) in clinical products, Int. J. Pharm. 495: 87-92, 2015.
- [2] J. Hadar, J. Garner, S. Skidmore, K. Park, H. Park, D. Kozak, and Y. Wang: Solvent-dependent PLGA solubility for separation of PLGAs with different lactide:glycolide ratios. 2018 Controlled Release Society (CRS) Annual Meeting (2018) Abstract 409.
- [3] S. Skidmore, J. Hadar, J. Garner, H. Park, K. Park, Y. Wang, and X. Jiang. Complex sameness: Separation of mixed poly (lactide-co-glycolide)s based on the lactide: glycolide ratio. *Journal of Controlled Release*, 300:174-184 (2019).
- [4] M. Zell, B.E. Padden, A.J. Paterick, K. Thakur, R.T. Kean, M.A. Hillmyer, and E.J. Munson. Unambiguous determination of the 13C and 1H NMR stereosequence assignments of polylactide using high-resolution solution NMR spectroscopy. *Macromolecules* 35 (20):7700-7707 (2002).

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